

Looking for Biological Monitoring Tools: Using Anemonia sulcata as Bioindicator Species to Assess Sunscreen and Ultraviolet Filter in Temperate Seas



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INTRODUCTION

The conservation of biodiversity and the social and economic drivers of marine ecosystems are directly linked to the identification of environmental stressors, the application of advanced marine monitoring techniques and the contribution to the value of marine resources.

Increasing pollution from contaminants of emerging concern, such as **sunscreens**, could accelerate the degradation of marine ecosystems and their resources.

Numerous studies associate the increased concentration of

In temperate latitudes, such as the **western Mediterranean**, pollution-related adverse effects such as bleaching of **cnidarians** (corals and anemones) have recently been documented.

In these areas, the symbiotic sea anemone *Anemonia sulcata* is gaining *increasing interest as a bioindicator species*, while at the same time the need for monitoring the bleaching phenomenon is increasing.

To assess the rate of adaptation and acclimatization in a changing climate, the defense mechanism of this potential bioindicator species against the stress of sunscreen pollution will be investigated in the laboratory.

The identification of potential **biomarkers** of this key species



UV filters with coral bleaching and reef deterioration in warm areas.

could significantly improve the capacity of environmental management in temperate marine areas.

Figure 1. Photo of Anemonia sulcata in the sea (https://www.malaga.es)

OBJECTIVE

To identify and characterize the pollution stress response to Benzophenone-3 (BP-3) and a sunscreen containing BP-3, at the individual and cellular levels in the temperate anemone specie Anemonia sulcata and its symbiont Symbiodinium sp., in order to identify biomarkers for monitoring emerging compound pollution in temperate seas.

Experimental design

Two essays of 7 days

BP-3

- Control
- Negative control (metanol)
- Range from 0,005 to 5000 µg/L of BP-3

Sunscreen contains BP-3 *

Clean hands control
5, 20, 50 and 100% of seawater matrix

* A seawater matrix was prepared by applying the recommended amount of sunscreen to hands according to the U.S. Food and Drug Administration (2 mg sunscreen/cm²) and immersing them in to 4 liters of seawater for 20 minutes (Danovaro et al., 2008).

Bleaching Measurement

The bleaching of the anemones was measured using a **Coral health chart** from the University of Queensland, Australia. (Danovaro et al., 2008). Example of bleaching. Anemone with 500 µg/L treatment of BP-3.





Bleaching Measurement



Status of Symbiodinium sp. (%)

Bleaching with BP-3



Bleaching with sunscreen



There is a causal relationship between the increase in BP-3 concentration and bleaching (p < 0,01).

It's a observed a positive correlation between DNA damage and GR (p<0.01) with increasing BP-3 concentration.</p>

Biomarkers of oxidative stress

- Significant differences in DNA damage and GR biomarkers (p<0.05) were determined between the control and environmental concentrations for the BP-3 assay.
- Tentacle tissues showed a more sensitive response to increasing BP-3 concentrations than column tissues in all what regards biomarker approach.
- No a clear tendance was observed between increasing percentage of sunscreen and biomarker responses.





The percentage of damaged *Symbiodinium* sp. showed that the percentage of damaged cells in the **column** increased with increasing concentration, reaching 61% at the highest concentration.

On the other hand, in the **tentacle**, the percentage of damaged cells remained at around 40% despite the increase in the concentration of BP-3.



(*) Significant differences with control (p < 0,05)

CONCLUSIONS

- ✓ BP-3 is potentially hazardous to marine ecosystems.
- ✓ BP-3 induce oxidative stress and provoke damage in symbiont organisms.
- BP-3 is an ingredient in sunscreens, and while we are aware of its presence in the environment, the mechanisms and timing of its transition to bioavailability for biota remain unknown, necessitating laboratory experiments.
- Biomarkers of oxidative stress and genotoxicity have shown to be potential monitoring tools for the assessment of contamination by BP-3.
- ✓ More research should be done regarding time and concentration testing of sunscreens

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 It is crucial to continue advancing the study of these compounds, both individually and within their original matrix, and to standardize protocols while facilitating the transfer of information to the industry.

Bibliography

Ani, C y Robson, B., 2021. Marine Pollution Bulletin, 166, 112223 Danovaro R., et al., 2008. Environmental Health Perspectives, 116, 441-447 Gardner, S. et al, 2017. BMC biology, 15, 1-15

Kibria, G. et al, 2021. Marine Pollution Bulletin, 167, 112364



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